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SAFETY AND EFFICACY OF UNWASHED FILTERED WOUND DRAINAGE BLOOD REINFUSED FOLLOWING ORTHOPAEDIC SURGERY

BY

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Our study demonstrates that the reinfusion of unwashed filtered postoperative drainage from orthopaedic wounds is an acceptable alternative to the transfusion of liquid preserved red blood cells. The volume of shed blood collected in the orthopaedic setting was about 350 ml, and no untoward effects on hemostasis were observed following reinfusion.

Safety and Efficacy of Unwashed Filtered Wound Drainage Blood Reinfused Following Orthopaedic Surgery

Autologous blood products are being used increasingly to fulfill the needs of patients undergoing orthopaedic and other surgical procedures. The major impetus for this practice is the concern about potential risks associated with homologous blood, e.g. the transmission of blood-borne infectious agents such as human immunodeficiency virus (HIV) and hepatitis viruses.

Autologous blood collected prior to surgery and shed blood collected intraoperatively and postoperatively are being used in place of homologous blood.1,2,3 Shed blood from orthopaedic wounds may be contaminated with fat particles, bone fragments, and methylmethacrylate monomer. Shed blood may also contain increased levels of plasma hemoglobin and fibrin degradation products as a result of hemolysis and clot lysis, and orthopaedic shed blood may also contain products of platelet and complement activation.1,2,3 Although shed blood is filtered before reinfusion, there are opposing views on whether or not washing is necessary.

This randomized, prospective clinical trial in 128 patients was undertaken to evaluate shed blood recovered after hip or knee replacement, or spinal fusion surgery. The effects of the reinfusion of shed blood collected

postoperatively by two autotransfusion systems were compared with the effects of the transfusion of liquid preserved autologous and homologous blood.

MATERIALS AND METHODS

one-hundred twenty-eight orthopaedic patients at four medical centers were randomized for study. Surgical procedures included total knee replacement, total hip replacement or spinal fusion. Written informed consent was obtained from each study subject. This was a randomized study of a) patients in whom shed blood was collected using one of two wound drainage systems, the Orth-evacTM or the SolcotransR, and reinfused postoperatively using the Pall 40 micron screen filter or the Pall RC100 polyester filter, and b) patients who did not receive shed blood but received autologous or homologous liquid preserved red blood cells. The patients who did not receive shed blood were treated with a standard wound drainage system (hemovac) and received homologous or autologous liquid preserved red blood cells.

Drainage catheters were placed in each patient using conventional surgical methods according to the manufacturer's directions. With the Solcotrans drainage system, forty (40) ml of acid citrate dextrose (ACD, NIH Formula A) was used; no anticoagulant was added with the Orth-evac drainage system.

Patients treated with the Orth-evac or Solcotrans were reinfused with at least 250 mls of shed blood within six hours after initiation of collection. The standards and

hospital policies of the American Association of Blood Banks (AABB) were observed for all transfusions. The need for transfusion of autologous or homologous liquid preserved red blood cells was determined by the clinical condition of the patient as assessed by the attending physician. In instances where less than 250 mls of shed blood was collected or a transfusion was not required, the patient was dropped from the study.

Blood samples were taken from the patient and from the collection device at various stages of the study to determine the patient's hematologic status, the characteristics of the blood in the collection chamber, and the effects of filtering on the blood. The following samples were collected from each patient: (1) baseline venous samples were drawn from patients prior to surgery (pre-surgery); (2) venous samples were drawn from patients prior to transfusion (post-surgery, pre-transfusion); (3) samples of shed blood were taken from the collection chamber and samples of autologous and homologous blood were collected from the blood bag prior to and following filtration; (4) venous blood samples were drawn from patients one and 24 hours post-transfusion. Samples were drawn into K3EDTA, NaCitrate and FDP tubes and were evaluated as follows: In the K3EDTA samples, plasma hemoglobin, complement C3a dys arg, hemoglobin, hematocrit, red blood cell count, white blood cell count, platelet count and fat content were measured; 4,5,6 In the NaCitrate samples, fibrinogen, plasminogen, antithrombin III, prothrombin time, partial thromboplastin time, thrombin time, Factor V, Factor VIII, D-dimers, antiplasmin, plasmin and thrombin were measured; and in FDP samples, fibrin degradation products were measured.7-12

In 24 patients at one medical center,
methylmethacrylate was measured in the shed blood five
minutes after insertion of the knee prosthesis and in the
patient one hour and 24 hours after reinfusion of the shed
blood.13,14 All patients were closely monitored through
measurements of blood pressure, heart rate, respiration
rate, and temperature, after surgery and after reinfusion of
the shed blood. Any adverse reactions or complications were
recorded.

DATA ANALYSIS: The means and standard deviations are reported. T-tests were used to compare data collected on the patients who received shed blood and the patients who received autologous and homologous liquid preserved red blood cells. Comparisons of data collected prior to and following the reinfusion of shed postoperative drainage were made using analysis of variance.15

RESULTS

All the patients in the study were of similar age, height, weight, and sex (Table 1), and types of surgical procedures to which the patients were subjected were similar (Table 2).

Nine of 84 patients from whom shed blood was collected were not reinfused and were excluded from the study: eight patients because the drainage collected was less than 250 ml, and one patient because the shed blood was stored at room temperature for longer than 6 hours.

The last 50 ml of shed blood collected was not reinfused because this portion of the shed blood is known to contain fat particles.

The patients who received 338 ml of shed blood collected in the Orth-evac system and the patients who received 336 ml of shed blood collected in the Solcotrans system also received autologous and homologous liquid-preserved red blood cells (Table 3, 4). Figure 1 reports the total volume of shed blood collected from each patient in the study.

Of 35 patients who received no shed blood, seven (7) were transfused with homologous liquid preserved blood and

eight (8) received both homologous and autologous red blood cells (Table 4). Six (6) of 54 patients (11%) who received shed blood postoperatively also were transfused with autologous and homologous liquid preserved red blood cells. Eight of 35 patients (22%) who did not receive shed blood were transfused with autologous and homologous liquid preserved red blood cells. The relative risk of transfusion with homologous blood was 0.4 in the patients who received shed blood group compared to patients who did not receive shed blood. The reinfusion of shed blood reduced the requirement for homologous blood by 60%. However, this reduction was not statistically significant (p=0.14) as measured by chi-square.

Studies of Blood Collected in the Transfusion Systems

Hematocrit (HCT), hemoglobin (HGB), and red blood cell counts (RBC) were lower in the post-operative shed blood than in the autologous and homologous liquid preserved red blood cells (Table 5). Platelet counts were somewhat lower in the shed blood than in the autologous or homologous liquid preserved red blood cells. Plasma hemoglobin levels were similar in shed blood samples collected in the OrthevacTM and the SolcotransR. The plasma hemoglobin level was lower in the postoperative shed blood than in the liquid stored red blood cells (Table 5).

Fibrinogen and antiplasmin levels were lower and Factor VIII and thrombin levels higher in the shed blood than in the autologous or homologous liquid preserved red blood cells (Table 5). Factor V levels were similar in shed blood and liquid preserved red blood cells (Table 5). Shed blood exhibited higher prothrombin times (PT) and thrombin times (TT) than the autologous or homologous liquid preserved red blood cells, but partial thromboplastin times (PTT) were similar (Table 5).

Plasminogen and plasmin levels were slightly higher in the shed blood than in the liquid preserved red blood cells (Table 5), but antithrombin III levels were similar (Table 5). Fibrin degradation products (FDP) and D-dimers were higher in the shed blood than in the liquid preserved red blood cells (Table 5). The shed blood exhibited higher concentrations of complement C3a dys arg (C3a) than the liquid preserved red blood cells (Table 5).

Methylmethacrylate monomer (MMA) was not detectable in the shed blood samples tested.

Greater concentration of fat particles (both <10 um diameter and 10-40 um diameter) were seen in the shed blood than in the liquid preserved red blood cells (Table 5). The RC100 filter appeared to be more effective than the 40 micron filter in removing fat particles and white blood cells (Table 6). The quality of the shed blood was not

significantly different whether it was filtered through the 40 micron Pall filter or the Pall RC100 polyester filter except for a lower concentration of fat particles and white blood cells in shed blood filtered through the RC100 polyester filter.

Systemic Blood Studies

systemic blood samples collected before surgery and at three points post-operatively revealed changes in hematologic parameters following surgery in all the patients studied whether they received shed blood or liquid preserved red blood cells mild reductions in hemoglobin concentrations and hematocrit values and increases in white blood cell counts were observed following surgery, and platelet counts were reduced for as long as 24 hours after transfusion.

At various post-surgical time points after the transfusion of the liquid preserved red blood cells, the level of fibrin degradation products (FDP) was roughly double that seen prior to surgery. In the patients who received the shed blood FDP was significantly increased one hour after transfusion to a level ten-fold that seen prior to surgery, but by 24 hours posttransfusion the levels were similar to those in the patients who received liquid preserved red blood cells (Table 7). D-dimer levels, which

also were increased at all post-surgical measurement times, were 3 to 4 times greater an hour after transfusion in the patients who received the shed blood than in the patients who received liquid preserved red blood cells, but similar 24 hours post-transfusion.

Fibrinogen levels were mildly to moderately decreased during surgery but within normal limits within 24 hours after transfusion of shed blood or liquid preserved red blood cells. Factor V, plasminogen, and antithrombin III levels were decreased within 24 hours after the transfusion of shed blood or liquid preserved red blood cells, but none of the other plasma proteins tested, including plasma hemoglobin, Factor VIII, antiplasmin, thrombin, plasmin, and complement C3a dys arg (C3a), showed any significant change, nor did prothrombin time (PT), partial thromboplastin time (PTT) or thrombin time (TT) (Table 7).

Methylmethacrylate monomer (MMA) was not detectable at any of the post-surgical time points (Table 7). In systemic blood samples, the levels of fat particles of both sizes (less than 10 um diameter (Fat <10) and between 10 and 40 um diameter (Fat 10-40)), were similar in the patients who received shed blood and those who received liquid preserved red blood cells (Table 7).

DISCUSSION

Our data indicate that reinfused unwashed orthopaedic wound drainage can be used safely and effectively in place of autologous or homologous liquid preserved red blood cells. When nonwashed, filtered shed blood was reinfused within six hours of collection, no febrile episodes or other transfusion reactions were observed.

Shed blood collected postoperatively using an autotransfusion device exhibited a lower hematocrit than autologous or homologous liquid preserved red blood cells. The shed blood had higher levels of activated complement C3a dys arg and prolonged prothrombin and thrombin time. Fibrin degradation products and D-dimers were significantly higher in the shed blood than in the liquid preserved red blood cells.

The decreases in fibrinogen and antiplasmin activity seen in the shed blood presumably were the result of clotting and lysis in the wound environment. The elevated thrombin level in the shed blood was probably due to conversion from prothrombin during the coagulation cascade. Factor VIII levels were also increased in shed blood.

The RC100 filter, designed to remove leukocytes, was more effective than the 40 um filter in removing both fat

particles and leukocytes from shed nonwashed blood. Neither filter affected any of the other measurements in shed blood.

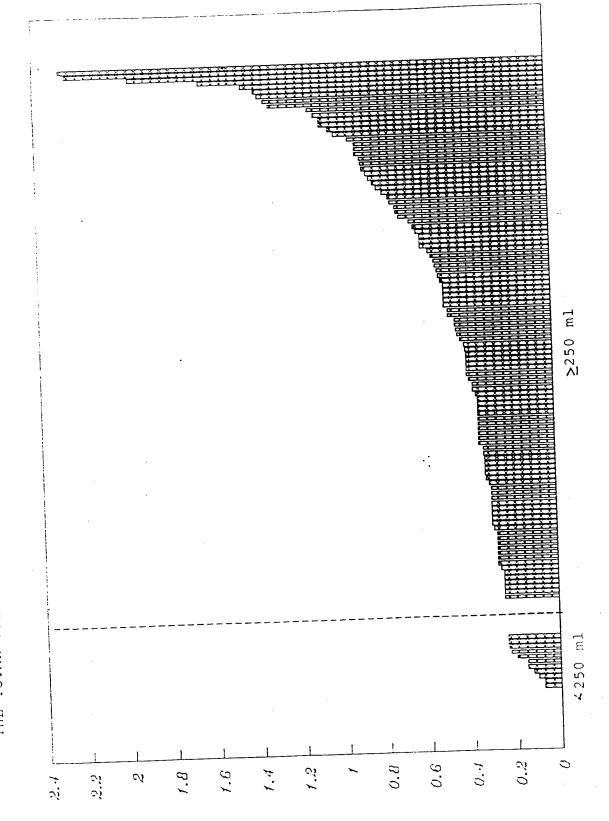
The results of systemic blood studies were not significantly different between the patients who received liquid preserved red blood cells and those reinfused with shed blood collected with either Orth-evac or Solcotrans system. This may have been due in part to the fact that only 350 ml of shed blood were reinfused. The main differences seen between the two patient groups were higher levels of fibrin degradation products and D-dimers in the patients who received the shed blood. Results of coagulation studies were similar in the two groups of patients. Neither methylmethacrylate monomer nor elevated fat particles were seen in systemic blood samples from either group, in spite of the fact that the shed blood was not washed before reinfusion.

The reinfusion of orthopaedic wound drainage has been steadily gaining acceptance. The reinfusion of wound drainage after elective knee arthroplasty reduced the demand for homologous banked blood by two-thirds.16 Patients reinfused with shed blood following cardiac surgery required only half as much homologous blood as patients who did not receive shed blood.17 Several investigators have reported on the successful use of reinfused shed blood following

cardiac surgery, with no evidence of septic, pulmonary, hepatic, or renal complication.18,19,20,21,22

Our study reported here demonstrates that the reinfusion of unwashed filtered postoperative drainage from orthopaedic wounds is an acceptable alternative to the transfusion of liquid preserved red blood cells. The volume of shed blood collected in the orthopaedic setting was about 350 ml, and no untoward effects on hemostasis were observed following reinfusion.

THE TOTAL VOLUME OF BLOOD COLLECTED FROM BACH PATIENT IN THE STUDY FIGURE 1



(LYONGGUGS)
LOLYT AOTHWE OF DRAINAGE COLLECTED (mil)

TABLE 1
DEMOGRAPHIC DATA

• '	ORTH-EVAC	SOLCOTRANS	CONTROL
Number of Patients Mean Age	44 67.9	40 66.3 54-82	44 62.5
Range Mean Height (in.)	41-82 65.7	67.2	67.3
Range Mean Weight (lb.)	58-72 178.2 109.5-276	59-73.5 190.7 104.5-260	190.4
Range Sex M F	109.3-270 18 26	20	23 21

TABLE 2
SURGICAL PROCEDURES

	ORTH-EVAC	SOLCOTRANS	CONTROL
		3.4	19
Total Knee	16	14	19
Bilateral Knees	16	10	7
Revision Knees	3	3	1
Total Hip	5	9	10
Revision Hips	4	3	7
Spine	0	1	0
TOTAL	44	40	44
Cemented	37	34	36
Uncemented	7	6	7
Cement Information	•		
Not Available	0	0	,1

TABLE 3

VOLUME OF SHED BLOOD COLLECTED AND REINFUSED

GROUP SOLCOTRANS	GROUP	<u>ORTH-EVAC</u>
Volume Collected (ml)		
Mean: SD: n: Range:	401 219 44 75- 930	466 276 40 125- 1370
Volume Reinfused (ml)		
Mean: SD: n: Range:	338 146 36 175- 750	366 201 39 200- 900

THE NUMBER OF UNITS OF AUTOLOGOUS AND HOMOLOGOUS LIQUID PRESERVED

TABLE 4

BLOOD THAT WERE COLLECTED AND TRANSFUSED INTO THE 3 DIFFERENT GROUPS OF PATIENTS

GROUP:	ORTH-EVAC	SOLCOTRANS	LIQUID PRESERVED
TOTAL # OF PATIENTS	36	39	43
TOTAL # OF PATIENTS PRE-DONATING BLOOD	26	28	35
# OF PREDONATED UNITS COLLECTED	60	80	92
% OF GROUP PREDONATE	ING 72	72	81
# OF PATIENTS RECEIVED PREDONATED BLOOD	7 IN G 22	24	30
# OF PREDONATED UNITS TRANSFUSED	47	58	70
% OF GROUP RECEIVING PREDONATED BLOOD	<i>G</i> 61	62	70
% OF PATIENTS WHO PREDONATED RECEIVING PREDONATED BLOOD	<i>G</i> 85	86	86
# OF PATIENTS RECEI HOMOLOGOUS BLOOD	VING 5	9	15
# OF HOMOLOGOUS UNITS TRANSFUSED	10	21	36
% OF GROUP RECEIVIN HOMOLOGOUS BLOOD	7G 14	23	34
# OF PATIENTS RECEI BOTH PREDONATED AND HOMOLOGOUS BLOOD	3	3	8
% OF GROUP RECEIVIN PREDONATED BLOOD AN HOMOLOGOUS BLOOD	IG BOTH ID 8	8	19

IN VITRO MEASUREMENTS OF POSTOPERATIVE ORTHOPEDIC DRAINAGE COLLECTED IN THE 2 RESERVOIRS AND LIQUID PRESERVED BLOOD BEFORE FILTRATION

TABLE 5

COLLECTION	POSTOPERATI	VE DRAINAGE	ANOVA*	LIQUID PRESERVED	SHED VS LIQUID PAIRED t-TEST
COLLECTION RESERVOIR:	ORTH-EVAC	SOLCOTRANS	"p" _	BLOOD	"p" .
		DODOCTICAL			
HEMATOCRIT (V%)	18 <u>+</u> 7	20 <u>+</u> 5	NS	52 <u>+</u> 27	<.0001
MEAN+SD	8	9		7	
N HEMOGLOBIN (qm/	_	•			
	6.0+2.4	6.7+1.6	NS	16.5 <u>+</u> 8.5	<.0001
MEAN±SD N	8	9		7	
RED BLOOD CELL	=	/mm3)			
MEAN+SD	2.04+0.73	2.10+0.60	NS	5.4 <u>+</u> 2.7	<.0001
MEAN_SD N	8	9		7	
WHITE BLOOD CEL		03/mm3)			
MEAN+SD	6.2+3.1	6.8+1.5	NS	1.8 <u>+</u> 2.1	<.001
N	8	9		7	
PLASMA HEMOGLO					
MEAN+SD	125+92	120+99	NS	197 <u>+</u> 390	NS
N N	27	25		20	
FIBRIN DEGRADAD	TION PRODUCT.	S(uq/ml)			
MEAN+SD	853 <u>+</u> 347	694 <u>+</u> 365	NS	11 <u>+</u> 2	<.0001
N	21	` 24		17	
FIBRINOGEN (mg)	/dl)				
MEAN+SD	23+14	35 <u>+</u> 31	NS	127 <u>+</u> 89	<.0001
N	20	18		18	
PLASMINOGEN (%)				
MEAN+SD	74 <u>+</u> 17	68 <u>+</u> 28	NS	56 <u>+</u> 31	<.01
N	21	23		19	
ANTITHROMBIN I	II (%)				
MEAN+SD	49 <u>+</u> 17	45<u>+</u>9	NS	50 <u>+</u> 35	NS
N	22	25		20	
PROTHROMBIN TI	ME (seconds)				
MEAN+SD	116 <u>+</u> 17	120 <u>+</u> 0	NS	66 <u>+</u> 48	<.0001
N	18	14		14	
ACTIVATED PART	IAL THROMBOP	LASTIN TIME (SE			<.02
MEAN+SD	120 <u>+</u> 0	120 <u>+</u> 0	NS	109 <u>+</u> 25	<.02
N	18	14		13	
THROMBIN_TIME	(seconds)			00:46	<.005
MEAN+SD	120 <u>+</u> 0	120 <u>+</u> 0	NS	92 <u>+</u> 46	~. 003
N	18	13		11	
FACTOR V CLOTT			27.0	12+4	<.01
MEAN+SD	10 <u>+</u> 0	10 <u>+</u> 0	NS	12 <u>+</u> 4 14	7.01
N	18	15		14	
FACTOR VIII CL		CIN (%)	NC	10.45	<.0001
MEAN+SD	48 <u>+</u> 23	35 <u>+</u> 28	NS	12 <u>+</u> 5 14	1,0001
N	17	14		74	

^{*}Analysis of variance: Orth-Evac versus Solcotrans Collection Reservoir

TABLE 5 (cont.)

COLLECTION	POSTOPERATIV	Æ DRAINAGE	ANOVA*	LIQUID PRESERVED	SHED VS LIQUID PAIRED T-TEST
RESERVOIR:	ORTH-EVAC	SOLCOTRANS	<u>"p"</u>	BLOOD	"p .
D-DIMER (ug/ml)					. 0001
MEAN+SD	>25000 <u>+</u> 0	>25000 <u>+</u> 0	NS	1166 <u>+</u> 1890	<.0001
- N	18	20		13	
ANTIPLASMIN (%	1				<.001
MEAN+SD	25 <u>+</u> 12	26 <u>+</u> 10	NS	46 <u>+</u> 27	<.001
N	17	16		15	
PLASMIN (%)				24.40	NS
MEAN+SD	90 <u>+</u> 52	78 <u>+</u> 39	NS	74 <u>+</u> 48	NS
N	7	8		7	
THROMBIN (IU/m.	<u>1)</u>		110	0.004+0.004	<.0001
MEAN+SD	0.185 <u>+</u> 0.080		NS	19	1.0001
N	21	25		13	
COMPLEMENT C3a		<u>ml)</u>		1442+944	<.0001
MEAN+SD	8140 <u>+</u> 6488	6851 <u>+</u> 3647	NS	1442 <u>+</u> 544 15	1.0001
N	25	21		15	
<u>METHYLMETHACRY</u>		(ug/m1)	NS	0.0 <u>+</u> 0.0	
MEAN <u>+</u> SD	0.0 <u>+</u> 0.0	0.0 <u>+</u> 0.0	NS	6	
N	8	10		Ü	
# FAT PARTICLE		. (10.400	NS	31+73	<.01
MEAN <u>+</u> SD	693 <u>+</u> 585	· 610 <u>+</u> 499	No	7	
N	8	9		,	
# FAT PARTICLE		22.22	NS	<u>0+</u> 0	
MEAN+SD	32 <u>+</u> 38	23 <u>+</u> 22 9	MP	7	
N	8	9		•	
# FAT PARTICLE		141	NS	0 <u>+</u> 0	
MEAN+SD	2 <u>+</u> 2	1 <u>+</u> 1	NO	7	
N	8	7		•	

^{*}Analysis of variance: Orth-Evac versus Solcotrans Collection Reservoir

IN VITRO MEASUREMENTS OF POSTOPERATIVE ORTHOPEDIC DRAINAGE AND LIQUID PRESERVED BLOOD PRIOR TO AND FOLLOWING FILTRATION

TABLE 6

	40u SC	REEN	POLYE				PRESERVED
COLLECTION		POST	PRE	POST	ANOVA*	PRE	
RESERVOIR:			FILT	ER	<u>"q"</u>	170u SC	CREEN FILTER
HEMATOCRIT (V							
MEAN+SD		24+9	20 <u>+</u> 7	22 <u>+</u> 9	NS	52 <u>+</u> 27	48+21
	10		7	7		7	7
HEMOGLOBIN (q.							
MEAN+SD	6.1+2.0	7.8 <u>+</u> 3.1	6.8 <u>+</u> 2.6	7.4 <u>+</u> 2.6	NS	16.5 <u>+</u> 8.5	15.4 <u>+</u> 6.8
		10	7	7		7	7
RED BLOOD CEL	L COUNT (x109/mm3)					
MEAN+SD	2.1 <u>+</u> 0.7	2.6 <u>+</u> 1.1	2.1 <u>+</u> 0.6	2.4 <u>+</u> 0.9	NS	5.4 <u>+</u> 2.7	5.0 <u>+</u> 2.1
N		10	7	7		7	7
WHITE BLOOD C	ELL COUNT	(x103/mm3)	-				
MEAN+SD	6.1 <u>+</u> 2.2	4.6 <u>+</u> 2.1	7.1 <u>+</u> 2.7	1.9 <u>+</u> 2.2	<0.05	1.8 <u>+</u> 2.1	1.4 <u>+</u> 1.8
N	10	10	7	7		7	/
PLASMA HEMOGL	OBIN (mg/	(dl)					100.100
MEAN+SD	125+106	136 <u>+</u> 124	120 <u>+</u> 85	115 <u>+</u> 85	NS	197 <u>+</u> 390	
N	25	25	27	27		20 .	19
FIBRIN DEGRAD	ATION PRO	DUCTS (ug/n	<u>11)</u>			22.0	11.2
MEAN+SD	748 <u>+</u> 365	668 <u>+</u> 388	792 <u>+</u> 364	853 <u>+</u> 423	NS	11 <u>+</u> 2	11 <u>+</u> 3 18
N	24	24	21	21		17	10
FIBRINOGEN (n	ng/dl	·				116:00	1201100
MEAN+SD	34 <u>+</u> 30	30 <u>+</u> 22		27 <u>+</u> 21	NS	116 <u>+</u> 90	138 <u>+</u> 100 17
N	19	21	19	18		20	17
PLASMINOGEN (<u>(&)</u>					E1 () 4	E6+30
MEAN+SD	69 <u>+</u> 20	71 <u>+</u> 21	73 <u>+</u> 27	76 <u>+</u> 38	NS	51 <u>+</u> 34	56 <u>+</u> 32 19
N	24	25	20	22		21	19
<u>ANTITHROMBIN</u>				40.44	NG	16136	48<u>+</u>3 8
MEAN+SD		-	49 <u>+</u> 16		NS	46 <u>+</u> 36 22	19
	25		22	21		22	10
PROTHROMBIN 3	TIME (seco	onds)		100.0	***	66 <u>+</u> 48	66+17
MEAN+SD	120 <u>+</u> 0	108 <u>+</u> 34			NS	14	14
N	4,	± ·	15	15		14	7-4
ACTIVATED PAR				onas)	NC	109+25	103 <u>+</u> 31
MEAN+SD		109 <u>+</u> 29			NS	13	13
N		16	15	15		13	10
THROMBIN TIME				100.0	NC	92 <u>+</u> 46	103 <u>+</u> 39
MEAN+SD	120 <u>+</u> 0		120 <u>+</u> 0	120 <u>+</u> 0	NS	11	12
N	17	15	14	14		* +	
FACTOR V CLO			10.0	10+0	NS	12 <u>+</u> 4	12 <u>+</u> 5
MEAN <u>+</u> SD	10 <u>+</u> 0	10 <u>+</u> 0	10 <u>+</u> 0	10 <u>+</u> 0	NO	14	12 <u>-</u> 3
N	18	18	15	15		7.4	**
FACTOR VIII				22±14	NS	12 <u>+</u> 5	15 <u>+</u> 8
MEAN+SD	45 <u>+</u> 32	44 <u>+</u> 34	39 <u>+</u> 17	33 <u>+</u> 14	NO	12 <u>-</u> 5 14	14
N	17	15	14	14		4	

^{*}Analysis of Variance: Pall RC100 Polyester versus Pall 40 Micron Screen Filter Post-Filtration

TABLE 6 (cont.)

	40u SCREEN	I	POLYESTE	R		LIQUID-PRESERVE	D CD
COLLECTION	PRE	POST	PRE	POST	ANOVA*	PRE F	POST
RESERVOIR:	FILTE	_	FILT	ER	<u>"p"</u>	170u SCREEN	FILTER
D-DIMER (ug/		<u></u>					
MEAN+SD	>25+0	>25+0	>25+0	>25+0	NS	0.66 <u>+</u> 0.49	0.52 <u>+</u> 0.39
mean <u>+</u> 3D N	21	22	17	17		13	11
- -							
ANTIPLASMIN	28+12	33 <u>+</u> 16	23 <u>+</u> 7	23+6	NS	42 <u>+</u> 28	45 <u>+</u> 27
MEAN <u>+</u> SD	20-12	18	13	12		19	15
N	20	16	13				
PLASMIN (%)	50.30	(2,21	97 <u>+</u> 49	103+41	NS	58±52	68+40
MEAN+SD	_	63 <u>+</u> 21	97 <u>+</u> 49	7	110	9	-8
N	8	8	1	,		-	
THROMBIN (IU	<u>/ml)</u>		0 00:0 00	0 21+0 10	NS	0.004 <u>+</u> 0.004	0.003+0.002
MEAN <u>+</u> SD			0.22 <u>+</u> 0.09	21	147	21	19
N	24	22	22	21		2.4	
COMPLEMENT C	3a dys Arq	(nq/ml)		0050.6345	NS	1442+944	2000+1289
MEAN <u>+</u> SD			8005 <u>+</u> 6816	9252+6345	NS	15	17
N	22	25	24	25		15	17
<u>METHYLMETHAC</u>	RYLATE MON	OMER (ug/m.	<u>1)</u>			0.040.0	0.0+0.0
MEAN+SD	0.0 <u>+</u> 0.0	0.0 <u>+</u> 0.0	0.0 <u>+</u> 0.0	0.0 <u>+</u> 0.0	NS	0.0 <u>+</u> 0.0	6
N	9	9	9	9		6	b
# FAT PARTIC	LES <10u/m	<u>1</u>					15.25
MEAN+SD	576 <u>+</u> 451	431 <u>+</u> 546	754 <u>+</u> 637		NS	31 <u>+</u> 73	15 <u>+</u> 35
– N	10	10	7	7		7	7
# FAT PARTIC	LES 10-40u	<u>/m1</u>					0.0
MEAN+SD		14 <u>+</u> 17	30 <u>+</u> 39	5 <u>+</u> 9	NS	0 <u>+</u> 0	0 <u>+</u> 0
N	10	10	7	7		7	7
# FAT PARTIC	LES >40u/m	<u>1</u>					0.0
MEAN+SD	2+2	1 <u>+</u> 2	1 <u>+</u> 2	0 <u>+</u> 1	NS	0 <u>+</u> 0	0 <u>+</u> 0
N	10	10	7	7		7	7

^{*}Analysis of Variance: Pall RC100 Polyester versus Pall 40 Micron Screen Filter Post Filtration

HEMATOLOGIC AND CLOTTING PROTEIN MEASUREMENTS IN PATIENTS PRIOR TO AND FOLLOWING THE REINFUSION OF POSTOPERATIVE SHED BLOOD COLLECTED INTO 2 DIFFERENT RESERVOIRS AND LIQUID PRESERVED BLOOD TABLE 7

	ORTH-EVAC COLLECTED	CLLECTED	6	801	SOLCOTRANS-COLLECTED POSTOPERATIVE DRAINA	DLLECTED S DRAINAGE			LIQUID PR	PRESERVED (CONTROL)	
og Rego	POSTOPERATIVE DRAINAGE POST-OP	E DRAINAG	3	PRE	POST-OP			€ð.	POST-OP		•
do OP	PRE TXN	1HR	24HR	do	PRE TXN	1HR	24HR	do	PRE TXN	IHR	24HR
II	7 7 6 6	7700	32+3	37+3	33+2	33+3	33+1	36±5	31±3	31±2	31+1
(EAN+SD 38±5 N 10	777 8 1) -1 & 2) វ - ស)		١٥	6	9	12	7	7	ហ
MOGLOBIN (qm/dl)	7		•	,	0	9 043 01	10 8±0.5	11.6+1.7		10.0+1.5 10	10.1+0.4
EAN+SD 12.3+1.7 10.6+1.3	10.6+1.3	10.8 ± 1.2 11.1 ± 1.3	11.1 ± 1.3	11.8±1.0	10.5±0.8	0. FI 0		12	7		ا س
N 10	ω	∞ .	v	0	n)				
	UNT (×109/r	nm3.1	r C	2 040 2	2 5+0 3	3.5+0.3	3.6+0.1	3.8+0.5	3.3+0.3	3.4+0.4	3.2±0.2
	4.2 ± 0.6 3.6 ± 0.4 3.6 ± 0.4	3.6±0.4	3./+0.5	υ - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1	-1 o	6	9	12	7		ហ
OT N		,	•								
TIE BLOOD CELL COUNT (XIUS) MINIS	12 043 3	2/ mm21 11/13+3.5	12.3+1.5	5.3+1.2	11.7+2.2	12.2±2.9	10.8±2.9	6.3±1.8	11.2±2.4	11.7±2.2	10.8+3.3
16AN+50 6.6+1.9 17.0±3.9 14.5±3.0 N 10 8 8	80.71	0.00	S	۱۵	6	6	9	12	7	7	ហ
Š	105/mm3)										101+20
AFAN+SD 294+75	241+49	227±52	238±34	262+68	223+65	194+64	195±42	257 <u>+</u> 63	7007 7	707/7	25.
N 10	۱۵	æ	ហ	æ	6	6	o	77	`)
SAN PLATELET VOLUME (x105/mm3)	UME (×105/	mm3.1		•	•		1111	0 0+0	9.0+0.7	9.6+1.3	8.4+0.5
MEAN+SD 8.6+1.0	8.6+1.0 8.6+1.0	8.7±1.0	7.7±0.5	9.1 ± 1.2	9.1±1.0	9.3±1.2	7 • 7 • 7 • 6	7.5.5.		7] ភេ
N 10	83		ហ	ω	א	ע	D	77	•		
JASMA HEMOGLOBIN (mg/dl)	(mg/d1)		V .	7 8+6 9	8,6+15,5	7.2+6.6	4.4+3.4	8.5+8.3	7.7±11.2	7.3±9.5	4.8+6.4
VEAN_SD 8.0+10.2 5.4+4.8	2 5.4 <u>+</u> 4.8	5.6 <u>+</u> 4./ 25	4.013.4	29	29		27	27	20	22	20
FGR.	N PRODUCTS	(uq/ml)				•		•	73+33	27+25	20+12
VEAN+SD 12+7	30+35	114+143	20±16	11+3	26±21	74+66	7077	7 TT	7 H C	61	17
N 21	19	20	2,1	24	24	23	6.3	0))	1	
-DIMER (ug/ml, x10 ²)	1 <u>-01</u> 3	•		6		20.4+6.4	4.4+2.6	0.6+0.2	4.6+5.7	5.8+4.3	2.4±1.5
MEAN+SD 1.0+0.7	7.9+7.2	19.046.9	4.9+5.6). OHO: 1	1	22	191	16	18	16	14
N 14	17	18	13	7.7		1	1				
IBRINOGEN (mg/dl)	7.		000	367036	286+63	289+88	433+122	341+91	307±101	288±110	
MEAN \pm SD 416 \pm 112 310 \pm 102 N 22 21	310 <u>±</u> 102 21	263 ± 66 19	440±190	24	22	22	21	20	20	20	16

TABLE 7 (cont.)

ORTH-EVAC COLLECTED POSTOPERATIVE DRAINAGE POST-OP	GE 24 H R	SO PRE	SOLCOTRANS-COLLECTED POSTOPERATIVE DRAINA POST-OP PRE TXN 1HR	SOLCOTRANS-COLLECTED POSTOPERATIVE DRAINAGE POST-OP PRE TXN 1HR	3E 24HR	PRE P	LIQUID PR BLOOD (C POST-OP PRE IXN	PRESERVED (CONTROL) 1HR	24HR
n	10	104±14	97 <u>+</u> 18 24	117 ± 20 22	83 <u>+</u> 13 20	105±9 20	90 <u>+</u> 18 20	93 ± 13	88 ± 23
91 <u>+</u> 18 20		104 <u>+</u> 15 26	90 <u>+</u> 13 25	90 <u>+</u> 13 25	86±15	110 <u>+</u> 11 20	91+14	92 ± 17 20	91 <u>+</u> 13 17
$12.0 \pm 1.5 \ 11.8 \pm 13 \ 1 $ $16 $	T *	11.1+1.2	11.5 ± 0.9 14	11.6 ± 1.0 13	11.7 ± 0.8 15	11.0+0.6 15	11.9 ± 1.4 14	13.5±3.5 15	12.3 ± 2.1 15
<u>TTIVATED PARTIAL THROMBOPLASTIN TIME (seconds)[£]</u> WEAN+SD 31.1+3.6 32.4+4.8 31.4+6.1 36.0+6.5 3 N 18 17 16 17	۲ ۴ 3	31.6 1 5.2	30.3 ± 3.8 14	29.9±3.6 14	34.3 ± 3.9	35.3 <u>+</u> 5.4	33.6 <u>+</u> 4.5 14	34.8 ± 7.2	35.0 <u>+</u> 8.0
10.1 ± 1.3 10	10	10.9 ± 2.2 13	12.5 ± 2.7 12	12.0 ± 2.3 11	10.6±1.7	10.5 ± 1.7 14	10.8 ± 2.1 12	11.3±2.2	10.1 ± 1.6 13
62 <u>+</u> 24 17	~	88 <u>+</u> 25 14	52 ± 17 14	47 <u>+</u> 5 15	57 ± 11 14	74 <u>+</u> 20 15	46 <u>+</u> 16 15	38 <u>+</u> 17 15	58 <u>+</u> 32 15
126 <u>+</u> 38 18	O.	97 <u>+</u> 25 14	119 <u>+</u> 57 14	120 <u>+</u> 47 13	102 + 28 13	92 <u>+</u> 32 14	101 <u>+</u> 41 15	96 <u>+</u> 42 15	109±51
75 <u>+</u> 16 19		83 <u>+</u> 14 16	72 ± 13	69 ± 18 16	81 <u>+</u> 16 16	83±9	77 ± 10 16	76 <u>+</u> 12 17	84 <u>+</u> 35
71+50	Σ.	56 <u>+</u> 15 7	69 <u>+</u> 36 8	59 <u>+</u> 23	58 <u>+</u> 19	58 <u>+</u> 15	60 ± 18	58 <u>+</u> 11 9	1 52 <u>+</u> 4 9 8
5+8 19		4+7 24	3 1 3 25	5 1 5 2 4	6 <u>+</u> 10 24	5±7 21	2 <u>+</u> 1 20	2 <u>+</u> 1 20	$\begin{array}{c} 3 \pm 4 \\ 19 \end{array}$

Excluding 8 patients with abnormal PT, aPTT and/or TT results: 4 patients who received liquid preserved red blood cells nly and 4 patients who received shed blood and liquid preserved red blood cells.

TABLE 7 (cont.)

ORTH-EVAC COLLECTED POSTOPERATIVE DRAINAGE	LE'CTED DRAINAGE	63	SOI P05	SOLCOTRANS-COLLECTED POSTOPERATIVE DRAINA	OLLECTED DRAINAGE			8 8	PRESERVED (CONTROL)	
PRE POST-OP	1HR	24HR	PRE OP	POST-OP PRE TXN	1HR	24HR	OP	PRE TXN	1HR	24HR
dys Arg (ng/ml) 23 591+584 6 27	05 <u>±</u> 510 21		504 <u>+</u> 314 22	690 <u>+</u> 876 22	890±1335 837±1108 20 20	837±1108 20	410 <u>+</u> 260 15	413 ± 303 15	429 <u>+</u> 295 16	471 <u>+</u> 201 16
THYLMETHACRYLATE MONOMER (uq/ml) (EAN+SD 0.0+0.0 0.0+0.0 0.0+0.0 N 9 9 2	0.0 <u>+</u> 0.0	0.0+0.0	1 1	0.0±0.0	0.0 ± 0.0 10	0.0+0.0	1 1	0.0+0.0	0.0±0.0	0.0+0.0
EAN_SD 4.1±8.7 7.3±18.8 3.6±9.6 0.0±0.0 8 10 8 5	9.6 <u>+</u> 9.	0.0±0.0	$0.6\frac{+1.3}{8}$	1.1 ± 2.5	4.4±11.5 5.7±12.2 '9 6	5.7 ± 12.2 6	0.6+1.5	0.0+0.0	1.1±1.7	0.0±0.0
FAT PARTICLES 10-40u/ml TEAN+SD 3.0+9.0 3.0+7.9 3 N 10 8	1.1 <u>+</u> 8.3 8	$3.1\pm 8.3 9.2\pm 18.4 0.0\pm 0.0$	0.0±0.0	0.4 ± 1.3	2.1 ± 6.0	3.3+7.5	0.1±0.3	0.0+0.0	0.0±0.0	0.0+0.0
FAT PARTICLES >40u/ml FEAN+SD 2.3+3.9 0.3+0.5 1.7+2.4 6.2+12.4 N 4 3 3 5	7+2.4	6.2 ± 12.4 5	0.0±0.0	0.1±0.3	0.0±0.0	0.0±0.0	0.0±0.0	0.0±0.0 0.0±0.0	0.0±0.0	0.0+0.0

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